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RESEARCH ARTICLE

FIELD PERFORMANCE OF SCHIMA WALLICHII (DC.) KORTH. SEEDLINGS INOCULATED WITH ECTOMYCORRHIZAL FUNGI

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ARTICLE INFO	ABSTRACT
Article History:	Symbiotic efficiency of ectomycorrhizal fungi on the growth and nutrient uptake of
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Symbiotic efficiency of ectomycorrhizal fungi on the growth and nutrient uptake of *Schima wallichii* seedlings were studied in two abandoned jhum land. Native ectomycorrhizal fungi (*Suillus bovinus, Boletus edulis* and *Scleroderma citrinum*) were inoculated using pure culture technique. Various growth characters viz. shoot height, leaf length, root length, root colar diameter and seedlings volume at various intervals responded significantly to all the fungal inoculants. A significant variation in phosphorus (P<0.01) content was observed between the treated and un-inoculated control. The formation and network of ectomycorrhizae appears to have an affirmative impact on the increased absorption of phosphorus resulting in better growth of the seedlings. Among the fungal inoculants, *Boletus edulis* successfully colonized and promoted the growth and nutrient uptake in *Schima wallichii* seedlings.

INTRODUCTION

Natural ecosystems depend on closed nutrient cycles to minimize losses of nutrients and optimize turnover within the soil-plant system. Any disturbance in the ecosystem generally disrupts these cycles resulting in rapid decline of plant productivity. The scale of human activity has become such that most of the ecosystems of the earth have been disrupted in some ways (Ehrlich, 1993). Deforestation and land degradation as a result of shifting cultivation (Jhum) is of great concern because of its impact on biodiversity and environmental issues. In Northeast India (with particular reference to Nagaland), jhum system of cultivation worked well when there was a balance between population and soil fertility as a result of longer fallow cycle. In recent years, the resilience of ecosystem has broken down and the land is increasingly deteriorating because of the increase in population, pressures on land, coupled with decreasing fallow cycles. In view of the unabated deforestation and decline in land productivity, plantation of native plant species has been encouraged to ameliorate the abandoned jhum lands. The chief advantage of using local tree species in reforestation programmes is that they are adapted to local environmental conditions, improve livelihood and economics of the local communities, who can utilize them and contribute to the conservation of biodiversity.

Degraded lands are generally poor in available nutrients limiting the growth of transplanted seedlings. Successful establishment of seedlings on such sites often fails without the presence of appropriate fungal partners (Kendrick, 2000; Lakhanpal, 2000). Ectomycorrhizal fungal inoculants provide the potential for improvement in quality and performance of out planted seedlings (Dominguez *et al.*, 2006; Quoreshi *et al.*, 2008; Rivero *et al.*, 2009). In nature, a large diversity of ectomycorrhizal fungi is available with varying capacity for colonizing plant roots and promoting growth. Ectomycorrhizal © Copy Right, IJRSR, 2014, Academic Journals. All rights reserved.

fungi can be easily isolated from the field especially from fungal fruiting bodies (Dodd and Thomson, 1994) and profitably launch with the existing reforestation programmes. Furthermore, host fungus specificity exists for many mycorrhizal fungi (Lesueur *et al.*, 2001; Wu *et al.* 2002; Cavallazzi *et al.*, 2007) which imply a need to select appropriate fungal isolates before embarking on inoculation for a particular host plant. Field trials of inoculated seedlings is an important stratagem as the full benefits of mycorrhiza can become evident only when plants are transplanted to the field and exposed to various stresses (Baum *et al.*, 2002; Perry etal., 1987; Quoreshi *et al.*, 2008).

MATERIALS AND METHODS

Study Site: The field trials were carried out in two abandoned jhum land:

Plot A: At Yimyu (1080.52m above msl), in the district of Mokokchung, Nagaland;

Plot B: At Lumami (962.25m above msl), in the district of Zunheboto, Nagaland.

Three native ectomycorrhizal fungi viz. *Suillus bovinus, Boletus edulis* and *Scleroderma citrinum* were isolated and multiplied on Modified Melin Norkran's medium (Marx, 1969). Seedlings of *Schima wallichii* were raised in nursery beds and transplanted in both the plots. One month after transplanting, the selected isolates of fungi were inoculated (5mm diameter block) near the roots of the seedlings, with each treatment replicating 25 times. A set of seedlings were maintained as control without any fungal inoculum. After confirming the symbiotic association, different growth characters of the seedlings per treatment were recorded for one year at an interval of four months. Seedlings volume was calculated as [(root colar - diameter)² x height or D²H].

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Percentage of mycorrhizal colonization/cm was calculated as follows (Sharma, 1981):

Ectomycorrhizae (%) =
$$\frac{\text{Total no. of mycorrhizal lateral}}{\text{Total No. of lateral rootlets}} \times 100$$

For the estimation of total phosphorus in plant tissue the wet tri-acid digestion procedure was followed as suggested by Allen (1974). Phosphorus was analyzed by molybdenum blue method (Jackson, 1973) and was calculated by the following formula:

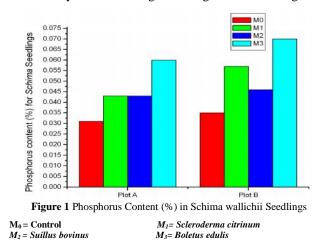
$$P(\%) = \frac{c (mg) x \text{ solution volume (ml)}}{10 x \text{ aliquot (ml) } x \text{ sample weight}} x 100$$

Where c = mg P obtained from the graph. The data was processed by analysis of variance (ANOVA).

RESULTS

All the fungal inoculants successfully established on *Schima wallichii* seedlings. The inoculated seedlings have superior growth response over seedlings not inoculated with ectomycorrhizal fungi (Table 1). There were statistically significant differences (Table 2) between the treated and control seedlings in terms of shoot height, leaf length (P<0.05), root length, root colar diameter and seedlings volume (P<0.01). Among the fungal inoculants, *Boletus edulis* proved superior for all the studied growth parameters in both the plots.

infected seedlings were able to generate numerous lateral roots of greater length, creating more sites and allowing a deeper penetration into soil where the unassociated roots cannot reach. The extensive development of the root system in the infected seedlings might have accounted for greater nutrient exploitation zone to the mycelium resulting in better growth of seedlings.



The capability of fungal inoculants to synthesize biologically active substances (Jackson *et al.*, 1964); production and supply of growth regulators (Heinrich *et al.*, 1989); and greater access to water (Stribley, 1987) can positively change the root physiology, growth,

	Plot A					Plot B			
Growth Parameters	M_4	M_8	M_{12}	M_{16}	M_4	M_8	M_{12}	Ν	M ₁₆
Shoot height (cm)	M_0	7.4	11.8	39.5	102.3	7.5	11.8	33.6	110.6
	M_1	7.7	13.6	37.9	112.3	7.9	13.8	38.6	130.2
	M_2	7.5	13.0	34.1	107.1	7.7	13.6	35.0	109.0
	M_3	7.7	13.9	38.0	129.0	7.8	13.9	43.2	136.5
	M_0	5.5	7.5	13.3	14.9	4.8	7.5	13.1	16.3
Lasflandth (and)	M_1	6.9	10.3	16.8	18.5	5.3	10.6	16.8	19.6
Leaf length (cm)	M_2	6.6	10.1	16.4	16.9	4.5	9.7	16.6	19.4
	M_3	6.8	10.4	16.5	19.5	5.3	10.7	16.9	20.0
	M_0	7.0	11.2	14.6	28.1	7.1	11.3	14.5	28.3
Root length (cm)	M_1	7.4	12.6	16.4	34.8	7.7	12.4	16.5	36.5
	M_2	7.3	12.4	16.4	35.5	7.4	12.5	16.4	36.5
	M_3	7.5	12.5	16.4	38.1	7.6	12.8	16.7	40.6
	M_0	0.16	0.21	0.26	0.38	0.16	0.20	0.26	0.39
Root colar	M_1	0.18	0.24	0.32	0.50	0.19	0.26	0.32	0.52
diameter (cm)	M_2	0.17	0.24	0.30	0.47	0.18	0.25	0.30	0.48
	M_3	0.19	0.24	0.34	0.53	0.19	0.26	0.34	0.55
	M_0	0.18	0.49	0.99	4.06	0.18	0.45	0.98	4.31
Seedlings	M_1	0.24	0.73	1.68	8.72	0.28	0.84	1.69	9.88
volume (cm ³)	M_2	0.21	0.71	1.46	7.86	0.23	0.78	1.48	8.42
	M_3	0.27	0.76	1.91	10.73	0.27	0.86	1.93	12.19

Table 1 Effect of ectomycorrhizal on the growth of S. wallichii seedlings

Each value is the mean of 5 replication.

 $M_0 = Control$

 $M_1 = Scleroderma\ citrinum$

M₂ = Suillus bovinus

 M_3 = Boletus edulis The uptake of phosphorus was greatly enhanced in seedlings colonized by ectomycorrhizal fungi (Figure 1). The results of ANOVA showed a significant variation in phosphorus (P<0.01) content between the treated and un-inoculated control (Table 3). Boletus edulis gave the best result with respect to

uptake of phosphorus by colonized seedlings in both the plots.

DISCUSSION

Ectomycorrhizal fungi are well known to boost the ability of plants to deal with various environmental stresses. The seedlings infected with ectomycorrhizal fungi showed a better growth response as compared to unassociated seedlings. The development and morphogenesis (Barker and Tagu, 2000; Chalot *et al.*, 2002; Luo *et al.*, 2009). Another benefit that seedling inoculation can bring is, the increased resistance against biotic (plant pathogen) and abiotic (presence of toxic elements) stresses (Chalot *et al.*, 2002; Garbaye 2000; Hall 2002; Smith and Read, 2008).

The root systems of ectomycorrhizal seedlings were capable of increased absorption of phosphorus, which could be only due to the formation and network of ectomycorrhizae and their ability to solubilize unavailable nutrients. The beneficial role of mycorrhiza with respect to the uptake of nutrients appears to be related to the nutrient depletion zone. Phosphorous is less soluble and relatively immobile in soil. The development and extent of extraradical hyphae is thought to play a major role in overcoming the P_i depletion zone (Plassard and Dell, 2010). The extraradical mycelium extends beyond the phosphorous depletion zone thereby increasing the uptake volume of the integrated root fungus system. The production of organic acids by ectomycorrhizal fungi which may release phosphorus from organic and inorganic forms by lowering the soil pH or chelation of metal ions (Malajczuk and Cromack, 1982) and; the ability to store and efficiently translocate phosphorus into the host (Grove and Le Tacon, 1993) may also contribute to the increased uptake of phosphorus.

Table 2 Analysis of variance (F) values of mycorrhizal
and non-mycorrhizal with various parameters.

	Source of variance	Variation between mycorrhizal and non - mycorrhizal	
	Shoot height	67.90*	
	Leaf length	67.60*	
Schima	Root length	8.4**	
wallichii	Root colar diameter	45.1**	
	Seedlings volume	26.1**	
= Significant at p <	< 0.05 level	**= Significant at $p < 0.01$ le	

Boletus edulis was the most efficient ectomycorrhizal fungi for *Schima wallichii* as it significantly increased the growth and uptake of nutrients by effectively colonizing the roots. The differential response of the host plant to different fungal inoculants could be attributed to the amount of ectomycorrhizae formed and the efficiency of mycobionts to improve the growth of host under varied ecological conditions (Browning and Whitney, 1993). The results are in agreement with the statement that ectomycorrhizal association in tree seedlings is far better than no such associations at all and; some species of ectomycorrhizal fungi have proven to be more beneficial to trees under certain environmental conditions than others (Agarwal and Sah, 2009).

Table 3 Analysis of variance (F) values of mycorrhizal and non-mycorrhizal with various parameters.

Analysis of Variance (F) Values of Mycorrhizal and Non- mycorrhizal with Various Parameters				
Schima wallichii	Source of Variance	Variation BetweenMycorrhizal and Non- mycorrhizal		
	Phosphorus	11.2**		
** = Significar	it at $p < 0.01$ level			

CONCLUSION

Schima wallichii is an important plant which finds many uses for the local people of the region. It produces good firewood and yields a medium-weight to heavy hardwood which is moderately durable and resistant to dry wood termites. The local people use the wood for construction that is under cover such as columns, beams, for flooring, interior fitting, panelling, door and window frames etc. The experimental inoculation of seedlings with ectomycorrhizal fungi like *Boletus edulis* holds a great promise for effective use of abandoned jhum lands. Large scale reforestation programmes can be attempted by using *Schima wallichii* seedlings which is of great economic value to the local population.

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References

- Agarwal, P. and Sah, P. 2009. Ecological Importance of Ectomycorrhizae in World Forest Ecosysyems. Nature and Science, 7(2): 107-116.
- Allen, S.E. 1974. Chemical analysis of ecological materials, Blackwell Scientific Publication, Oxford.
- Barker, S.J. and Tagu, D. 2000. The roles of auxins and cytokinins in mycorrhizal symbiosis. J. Plant Growth Regul., 19: 144-154.
- Baum, C., Stetter, U. and Makeschin, F. 2002. Growth response of *Populus trichocarpa* to inoculation by the ectomycorrhizal fungus *Laccaria laccata* in a pot and a field experiment. For. Ecol. Manag., 163: 1-8.
- Browning, M.H.R. and Whitney, R.D. 1993. Infection of containerized jack pine and black spruce by *Laccaria* species and *Thelephora terrestris* and seedlings survival and growth after outplanting. Can. J. For. Res., 23:330-333.
- Cavallazzi, J.R.P., Filho, O.K., Stu^{*}rmer, S.L., Rygiewicz, P.T. and de Mendonça, M.M. 2007. Screening and selecting arbuscular mycorrhizal fungi for inoculating micropropagated apple rootstocks in acid soils. Plant Cell, Tissue and Organ Culture, 90: 117-129.
- Chalot, M., Javelle, A., Blaudez, D., Lambilliote, R., Cooke, R., Sentenac, H., Wipf, D. and Botton, B. 2002. An update on nutrient transport processes in ectomycorrhizas. Plant Soil, 244: 165-175.
- Dodd, J.C. and Thomson, B.D. 1994. The screening and selection of inoculants arbuscular-mycorrhizal and ectomycorrhizal fungi. Plant and Soil, 159: 149-158.
- Dominguez, J.A., Rodriguez Barreal J.A. and Saiz de Omeñaca, J.A. 2006. The influence of mycorrhization with *Tuber melanosporum* in the afforestation of a Mediterranean site with *Quercus ilex* and *Quercus faginea*. For. Ecol. Manag., 231: 226-233.
- Ehrlich, P.R. 1993. The scale of the human enterprise. In: Saunders DA, Hobbs RJ, Ehrlich PR (eds) Nature conservation 3: Reconstruction of fragmented ecosystemglobal and regional perspectives: Surrey Beatty & Sons, Chipping Norton, New South Wales, pp.3-8.
- Garbaye, J. 2000. The role of ectomycorrhizal symbiosis in the resistance of forest to water stress. Outlook Agric., 29: 63-69.
- Grove, T.S., LeTacon, F. 1993. Mycorrhiza in plantation forestry. In: Ingram DS, Williams PH (eds) Advances in Plant Pathology: Academic Press, New York, USA, pp. 191-227.
- Hall, J.L. 2002. Cellular mechanisms for heavy metal detoxification and tolerance. J. Exp. Bot., 53: 1-11.
- Heinrich, P.A., Muligen, D.R. and Patrick, J.F. 1989. The effect of ectomycorrhizal on the phosphorus and dry weight acquisition of Eucalyptus seedlings. Plant and Soil, 108: 147-149.
- Jackson, M.L. 1973. Soil Chemical Analysis, 2nd medium reprint, Prentice Hall of India, New Delhi.
- Kendrick, B. 2000. The fifth kingdom. Third edition. Focus Publishing, Newburyport.

- Lakhanpal, T.N. 2000. Ectomycorrhiza an overview. In: Mukerji KG, Chamola BP, Singh J (eds) Mycorrhizal biology: Kluwer Academic / Plenum Publishers, New York, pp. 101-118.
- Lesueur, D., Ingleby, K., Odee, D., Chamberlain, J., Wilson, J., Manga, T. T., Sarrailh, J. M. and Pottinger, A. 2001. Improvement of forage production in *Calliandra calothyrsus*: methodology for the identification of an effective inoculum containing *Rhizobium* strains and arbuscular mycorrhizal isolates. Journal of Biotechnology, 91: 269-282.
- Luo, Z.B., Janz D., Jiang, X., Göbel, C., Wildhagen, H., Tan, Y., Rennenberg, H., Feussner, I. and Polle, A. 2009. Upgrading root physiology for stress tolerance by ectomycorrhizas: insights from metabolite and transcriptional profiling into reprogramming for stress anticipation. Plant Physiol., 151: 1902-1917.
- Malajczuk, N. and Cromack, J.K. 1982. Accumulation of calcium oxalate in the mantle of ectomycorrhizal roots of *Pinus radiate* and *Eucalyptus marginata*. New Phytologist, 92: 527-531.
- Marx, D.H. 1969. Antagonism of mycorrhizal fungi to root pathogenic fungi and soil bacteria. Phytopathology, 59: 153-163.

- Perry, A.D., Molina, R. and Amaranthus, P.M. 1987. Mycorrhizae, mycorhizospheres, and reforestation: Current knowledge and research needs. Can. J. For. Res., 17: 929-940.
- Plassard, C. and Dell, B. 2010. Phosphorus nutrition of mycorrhizal tress. Tree Physiology, 30: 1129-1139.
- Quoreshi, A.M., Piche, Y. and Khasa, D. P. 2008. Field performance of conifer and hardwood species 5 years after nursery inoculation in the Canadian Prairie Provinces. New. For., 35: 235-253.
- Rivero, S.H.T., Moorillon, V.G.N. and Borunda, E.O. 2009. Growth, yield, and nutrient status of pecans fertilized with biosolids and inoculated with rhizosphere fungi. Bioresour Technol., 100: 1992-1998.
- Sharma, G.D. 1981. Ecological studies on mycorrhizae of pine (*Pinus kesiya*). Ph.D Thesis submitted to the North Eastern Hill Univ. Shillong.
- Smith, S.E. and Read, D.J. 2008. Mycorrhizal Symbiosis, 3rd edn. Academic, London.
- Stribley, D.P. 1987. Ecophysiology of VA mycorrhizal Plants. In: Safir GR (ed) Mineral nutrition: CRC Press Boca Raton, FL, pp.59-70.
- Wu, T., Hao, W., Lin, X. and Shi, Y. 2002. Screening of arbuscular mycorrhizal fungi for the revegetation of eroded red soils in subtropical China. Plant and Soil, 239: 225-235.
