



ISSN: 0976-3031

Available Online at <http://www.recentscientific.com>

International Journal of Recent Scientific Research  
Vol. 6, Issue, 6, pp.4571-4575, June, 2015

International Journal  
of Recent Scientific  
Research

## RESEARCH ARTICLE

# ANTIBACTERIAL ACTIVITY OF A BIFUNCTIONALIZED ALLENE ETHANOL EXTRACTS

Ignatova-Ivanova TS<sup>1\*</sup>, Ivelina Stefanova<sup>1</sup>, Ismail E. Ismailov<sup>2</sup>, Ivaylo K. Ivanov<sup>2</sup>  
and Valerij Ch. Christov<sup>2</sup>

Faculty of Natural Sciences, Konstantin Preslavsky University of Shumen, 115, Universitetska Str.,  
BG-9712 Shumen, Bulgaria

### ARTICLE INFO

#### Article History:

Received 2<sup>nd</sup>, May, 2015  
Received in revised form 10<sup>th</sup>,  
May, 2015  
Accepted 4<sup>th</sup>, June, 2015  
Published online 28<sup>th</sup>,  
June, 2015

#### Key words:

**BA-2** (Dimethyl 1-(1-hydroxyethyl)-3-methylpenta-1,2-dienephosphonate), antibacterial activity, antibiotic.

### ABSTRACT

**Background and Purpose:** Antibacterial effects of a Bifunctionalized Allene with unprotected hydroxy group (Dimethyl 1-(1-hydroxyethyl)-3-methylpenta-1,2-dienephosphonate) (**BA-2**) on pathogenic Gram-positives and Gram-negatives bacteria had been established. **BA-2** (50mg/ml, 25mg/ml, 12.5mg/ml, 6.25mg/ml and 3.125mg/ml) exerted different inhibitory effect on different bacterial cells *in vitro*. The effects of **BA-2** on prokaryotic cells have not been studied. The present study was aimed to assess the antibacterial activity of **BA-2** on pathogenic Gram-positive and Gram-negative bacteria.

**Experimental approach:** *In vitro* antimicrobial test: *Escherichia coli* 3398, *Staphylococcus aureus* 745, *Bacillus subtilis* 6633, *Salmonella Typhimurium* 3591, *Listeria monocytogen* 863 and *Enterobacter aerogenes* 3691 were treated for 24 hours with **BA-2** (50mg/ml, 25mg/ml, 12.5mg/ml, 6.25mg/ml and 3.125mg/ml), Sefpotec (250 mg/ml). The antibacterial activity was assayed by the well diffusion method with digital caliper. **Determination of minimum inhibitory concentrations (MICs):** The MIC of **BA-2**, that shows antimicrobial activity, were determined by methods as described by [16] and MICs were read in µg/ml after overnight incubation at 37°C. All experiments were made in replicate. **Determination of Minimum bacteriocidal concentration (MBC):** The MBC were carried out to check whether the test microbes were killed or only their growth was inhibited. Nutrient Agar was prepared and sterilized at 121°C for 15 minutes, the medium was poured into sterile petridishes and were allowed to cool and solidify. The contents of the MIC in the serial dilution were then subcultured onto the prepared medium, incubation was made at 37°C for 24 h, after which each plate was observed for colony growth. The lowest concentration of the **BA-2** without a colony growth was recorded as the MBC. **BA-2** had higher antibacterial activity than tested antibiotic – Sefpotec.

**Key Results:** The results revealed variability in the inhibitory concentrations of **BA-2** for given bacteria. **BA-2** at concentration 50 mg/ml for 24 hours notably inhibited growth of *aerogenes* 3691 (25.52 mm mean zone of inhibition), *E. coli* 3398 (30.48 mm mean zone of inhibition) and *S. Typhimurium* 745 (21.22 mm mean zone of inhibition). On the contrary, **BA-2** had no activity against *L. monocytogen* 863 (17.39 mm mean zone of inhibition), which are comparable to the inhibitory effect of standard drug. **BA-2** did not inhibit *S. aureus* 745 and *B. subtilis* 6633.

**Conclusions and Implications:** Based on the results obtained we can conclude that the examined **BA-2** has bactericidal activity towards pathogenic bacteria, but in different concentrations.

The **BA-2** possesses biological activity, which is not well studied. We know only from literary data that they are used for inhibiting the biosynthesis of sterol from the pathogen responsible for *Pneumocystis carinii* pneumonia (PCP) – a disease similar to AIDS [1]. In our previous studies was shown that the Bifunctionalized Allene with protected hydroxy group (Dimethyl 3-methyl-1-[1-(tetrahydro-2H-pyran-2-yloxy)-ethyl]-hepta-1,2-dienephosphonate) (**BA-1**) exhibited antibacterial [10] and antifungal activity [11]. The results obtained show for the first time the existence of antibacterial activity of **BA-2** towards various pathogenic bacteria.

**Copyright © Ignatova-Ivanova TS et al** This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original work is properly cited.

### INTRODUCTION

The decreasing effectiveness of antibiotics in treating common infections has quickened in recent years, and with the arrival of untreatable strains of carbapenem-resistant Enterobacteriaceae, we are at the dawn of a postantibiotic era [7].

In high-income countries, continued high rates of antibiotic use in hospitals, the community, and agriculture have contributed to selection pressure that has sustained resistant strains [14], forcing a shift to more expensive and more broad-spectrum antibiotics. In low income and middle-income countries (LMICs), antibiotic use is increasing with rising incomes, high

\*Corresponding author: Ignatova-Ivanova TS

Faculty of Natural Sciences, Konstantin Preslavsky University of Shumen, 115, Universitetska Str., BG-9712 Shumen, Bulgaria

rates of hospitalisation, and high prevalence of hospital infections. Resistance arises as a consequence of mutations in microbes and selection pressure from antibiotic use that provides a competitive advantage for mutated strains.

Suboptimum antibiotic doses help stepwise selection of resistance. Resistance genes are borne on chromosomal, and increasingly, on transmissible extrachromosomal elements. The resulting resistant clones—eg, methicillin-resistant *Staphylococcus aureus* (MRSA) USA 300, *Escherichia coli* ST131, and *Klebsiella* ST258) are disseminated rapidly worldwide. This spread is facilitated by inter-species gene transmission, poor sanitation and hygiene in communities and hospitals, and the increasing frequency of global, travel, trade, and disease transmission [7,14,17]. The lack of understanding of the unique features and risk of resistance has paved the way for the present epidemic. Countries that have implemented comprehensive national strategies have been the most successful in controlling resistance [6,18,19,20]. These strategies include, but are not restricted to, good health-care infrastructure and health insurance for all; limited drug advertising; surveillance of antibiotic use and to detect resistance in human beings and animals; policies for prudent antibiotic use in human beings and animals; standardized infection control policies and sufficient staffing; antibiotic stewardship programmes in hospitals and other health-care facilities; and isolation or decontamination of patients with resistant organisms [20].

In this paper, the antibacterial activity of a Bifunctionalized Allene with unprotected hydroxy group (*Dimethyl 1-(1-hydroxyethyl)-3-methylpenta-1,2-dienephosphonate*) (**BA-2**) has been studied as part of the exploration for new and novel bio-active compounds.

## MATERIALS AND METHODS

### Test organisms

*Escherichia coli* 3398, *Staphylococcus aureus* 745, *Bacillus subtilis* 6633, *Salmonella Typhimurium* 3591, *Listeria monocytogenes* 863 and *Enterobacter aerogenes* 3691 were obtained from the Collection of the Department of General and Applied Microbiology, Sofia University. All the isolates were checked for purity and maintained in slants of Nutrient agar.

### Media used

Nutrient Agar (Bio life 272-20128, Milano, Italia) was the medium used as the growth medium for the microbes.

### Compound tested

Bifunctionalized Allene with unprotected hydroxy group (*Dimethyl 1-(1-hydroxyethyl)-3-methylpenta-1,2-dienephosphonate*) (**BA-2**) was synthesised in the Laboratory of Toxicological Chemistry, Department of Organic Chemistry & Technology of the Konstantin Preslavsky University of Shumen, Bulgaria (figure 1) [12].

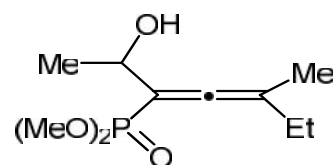


Figure 1 Structural formula of BA-2

*Dimethyl 1-(1-hydroxyethyl)-3-methylpenta-1,2-dienephosphonate* (**BA-2**). Yellow oil, yield: 80%. Rf0.58; IR (neat, cm<sup>-1</sup>): 1254 (P=O), 1956 (C=C=C), 3372 (OH). <sup>1</sup>H-NMR (600.1 MHz): 0.98 (t, *J* = 7.5 Hz, 3H, Me-CH<sub>2</sub>), 1.42 (dd, *J* = 6.1 Hz, *J* = 10.2 Hz, 3H, Me-CHO), 1.78 (d, *J* = 6.6 Hz, 3H, Me-C=), 2.02–2.10 (m, 2H, Me-CH<sub>2</sub>), 2.70 (s, 1H, OH), 3.78 (d, *J* = 11.6 Hz, 3H, MeO), 4.67–4.72 (m, 1H, Me-CHO). <sup>13</sup>C-NMR (150.9 MHz) = 12.2 (*J* = 7.6 Hz), 18.4 (*J* = 6.4 Hz), 23.2 (*J* = 7.5 Hz), 27.4 (*J* = 9.2 Hz), 52.6 (*J* = 6.2 Hz), 66.9 (*J* = 10.3 Hz), 96.3 (*J* = 192.3 Hz), 104.4 (*J* = 15.9 Hz), 208.9 (*J* = 5.4 Hz). <sup>31</sup>P-NMR (242.9 MHz): 21.1. Anal. Calcd for C<sub>10</sub>H<sub>19</sub>O<sub>4</sub>P (234.23): C 51.28, H 8.18. Found: C 51.21, H 8.13.

### Preparing the solution of BA-2

The solutions of **BA-2** (50mg/ml, 25mg/ml, 12.5mg/ml, 6.25mg/ml and 3.125mg/ml) were freshly prepared in ethanol.

### Assay for Antifungal Activity

Antimicrobial assay was performed by the well diffusion method using soft 0.8% agar. Agar medium was added to sterile Petri dishes seeded with 100 μl of each test bacterial strains. Wells of equal distance were dug on the seeded plates. Each well was filled up with 100 μl of the **BA-2** and antibiotics tested. After adjusting the pH at 6.5 by NaOH, the activity of the **BA-2** was checked. The plates were incubated at 37°C for 48 hours. The antibacterial activity was assayed by measuring the diameter of the inhibition zone formed around the well [4]. All experiments were performed in triplicate.

### Determination of Minimum inhibitory concentrations (MICs)

The minimum inhibitory concentrations of **BA-2**, that shows antimicrobial activity, were determined by 2-fold dilution methods as described by [16] and MICs were read in μg/ml after overnight incubation at 37°C. All experiments were made in replicate.

### Determination of Minimum fungal concentration (MBC)

The MBC were carried out to check whether the test microbes were killed or only their growth was inhibited. Nutrient Agar was prepared and sterilized at 121°C for 15 minutes, the medium was poured into sterile petri dishes and were allowed to cool and solidify. The contents of the MIC in the serial dilution were then sub cultured onto the prepared medium, incubation was made at 37°C for 24 h, after which each plate was observed for colony growth. The lowest concentration of the **BA-2** without a colony growth was recorded as the MBC.

## RESULTS

In the present study the effects of **BA-2** on six pathogenic Gram-positive and Gram-negative bacteria were evaluated. The effects were compared with widely used antibiotic Sefpotec. According to NCCLS, the antibiotic Sefpotec used is known to have broad spectrum antibacterial activity [15]. The effects of **BA-2** on the microorganisms were summarized in Table 1.

**Table 1** Effect of **BA-2** on test organisms.

Microorganisms	Zone of inhibition (mm) <sup>a</sup>
<i>S. aureus</i> 745 Gram- positive	-
<i>. aerogenes</i> 3691 Gram-negative	25.52±0.12
<i>E. coli</i> 3398 Gram-negative	30.48±0.02
<i>B. subtilis</i> 6633 Gram- positive	-
<i>L. monocytogen</i> 863 Gram- positive	17.39±0.10
<i>S. Typhymurium</i> 745 Gram-negative	21.22±0.17
Ethanol(96%) (Negative control)	11.20±0.19
Sefpotec 250µg/ml	21.10±0.05

<sup>a</sup>Data are presented as average values ± standard deviation in mm.

**BA-2** at concentration 50 mg/ml for 24 hours notably inhibited growth of *. aerogenes* 3691 (25.52mm mean zone of inhibition), *E. coli* 3398 (30.48mm mean zone of inhibition) and *S. Typhymurium* 745 (21.22 mm mean zone of inhibition). On the contrary, **BA-2** had no activity against *L. monocytogen* 863 (17.39mm mean zone of inhibition), which are comparable to the inhibitory effect of standard drug. **BA-2** did not inhibit *S. aureus* 745 and *B. subtilis* 6633.

Our assay for antibacterial activity of **BA-2** was conducted by testing different concentrations of the compound on various pathogens to determine the MICs. We used five concentrations – 50mg/ml, 25mg/ml, 12.5mg/ml, 6.25mg/ml and 3.125mg/ml. The results are shown in Table 2.

**Table 2** The MIC of **BA-2**

Microorganisms	MIC (mg/ml) <sup>a</sup>				
	50mg/ml	25mg/ml	12.5mg/ml	6.25mg/ml	3.125mg/ml
<i>S. aureus</i> 745	-	-	-	-	-
<i>. aerogenes</i> 3691			+		
<i>E. coli</i> 3398			+		
<i>B. subtilis</i> 6633	-	-	-	-	-
<i>L. monocytogen</i> 863		+			
<i>Salmonella Typhymurium</i> 745		+			

<sup>a</sup>Results are mean ± SEM of three separate trails.

**Table 3** The MBC of **BA-2**

Microorganisms	MBC (mg/ml) <sup>a</sup>				
	50mg/ml	25mg/ml	12.5mg/ml	6.25mg/ml	3.125mg/ml
<i>S. aureus</i> 745	-	-	-	-	-
<i>. aerogenes</i> 3691	+				
<i>E. coli</i> 3398	+				
<i>B. subtilis</i> 6633	-	-	-	-	-
<i>L. monocytogen</i> 863		+			
<i>Salmonella Typhymurium</i> 745	+				

<sup>a</sup>Results are mean ± SEM of three separate trails

The results revealed variability in the inhibitory concentrations of **BA-2** for given bacteria. MIC of **BA-2** at concentration 25 mg/ml for 24 hours notably inhibited growth of *L. monocytogen* 863 and *Salmonella Typhymurium* 745. In contrast, MIC of **BA-2** at concentration 12.5 mg/ml for 24 hours notably inhibited growth only of Gram-negative bacteria *. aerogenes* 3691 and *E. coli* 3398. Our next task was to determine the Minimum bactericidal concentration (MBC) in regards with determining the bactericidal or bacteriostatic

activity of the examined **BA-2**. We used five concentrations – 50mg/ml, 25mg/ml, 12.5mg/ml, 6.25mg/ml and 3.125mg/ml. The results are shown in Table 3.

MBC of **BA-2** at concentration 50 mg/ml for 24 hours notably inhibited growth only Gram-negative bacteria of *Salmonella Typhymurium* 745, *. aerogenes* 3691 and *E. coli* 3398. For *S. aureus* 745 and *B. subtilis* 6633 MBC it was not reported. MBC of **BA-2** at concentration 25 mg/ml for 24 hours notably inhibited growth only Gram-positive bacteria of *L. monocytogen* 863.

Based on the results obtained we can conclude that the examined **BA-2** has bactericidal activity towards both pathogenic bacteria, but in different concentrations.

The **BA-2** possesses biological activity, which is not well studied. We know only from literary data that they are used for inhibiting the biosynthesis of sterol from the pathogen responsible for *Pneumocystis-carinii* pneumonia (PCP) - a disease similar to AIDS [16]. In our previous studies was shown that the Bifunctionalized Allen with protected hydroxy group (*Dimethyl 3-methyl-1-[1-(tetrahydro-2H-pyran-2-yloxy)-ethyl]-hepta-1,2-dienephosphonate*) (**BA-1**) exhibited antibacterial [10] and antifungal activity [11]. The results obtained show for the first time the existence of antifungal activity of **BA-2** towards various pathogenic yeast and fungi. Even before penicillin was introduced, resistant strains of bacteria had been detected [1-3]. The selection pressure caused by the use of millions of tonnes of antibiotics over the past 75 years since antibiotics were introduced has made almost all disease-causing bacteria resistant to antibiotics commonly used to treat them.

The rapid evolution of bacterial resistance is clear in the case of  $\beta$ -lactamases class of antibiotics. Nearly 1000 resistance-related  $\beta$ -lactamases that inactivate these antibiotics have been identified, a tenfold increase since before 1990 [8]. Antibiotic use is a main driver of selection pressure that contributes to resistance, and because consumers do not understand this problem, the drugs are among the world's most commonly purchased. In high-income countries, patients with resistant infections can turn to more expensive, newer-generation

antibiotics, but in developing countries, where infectious diseases are common and the burden is high, patients might be unable to obtain or to afford second-line treatments. Large differences in the frequency of resistant infections have been noted, both across European countries [9] and among regions of the USA [5,13].

Bacteria which cause disease react to the antibiotics used as treatment by becoming resistant to them, sooner or later. This natural process of adaptation, antimicrobial resistance, means that the effective lifespan of antibiotics is limited. Unnecessary use and inappropriate use of antibiotics favours the emergence and spread of resistant bacteria. A crisis has been building up over decades, so that today many common and life-threatening infections are becoming difficult or even impossible to treat, sometimes turning a common infection into a life-threatening one. It is time to take much stronger action worldwide to avert a situation that entails an ever increasing health and economic burden [20]. Considering antibiotics resistance of most Gram-positive and Gram-negative bacteria, these Bifunctionalized Allene offer good and promising prospects for the treatment of diseases caused by these. The occurrence of drug resistant strains with less susceptibility to antibiotics due to mutation challenges the researchers to invent newer drugs. At this scenario, evaluation of antimicrobial substances from various sources is considered to be a pivotal role. Nevertheless, further studies are required to explore the mechanism of biochemical active principle in the Bifunctionalized Allenes for the inhibitory action on various pathogens selected in the study.

## CONCLUSION

The Bifunctionalized Allene with unprotected hydroxy group **BA-2** at 50mg/ml, 25mg/ml, 12.5mg/ml, 6.25mg/ml and 3.125 mg/ml concentrations showed significant anti bacterial activity on selected pathogens in clinical isolates.

## Acknowledgement

Support from the Research Fund of the Konstantin Preslavsky University of Shumen (Project No. RD-08-213/10. 03. 2014, Department of Biology) and (Projects Nos. RD-08-208/08. 02.2014 and RD-08-248/06. 03.2015, Department of Organic Chemistry & Technology) and Human Resources Development Operational Programme of the European Union (BG051PO001-3.3.06-0003/2012) is acknowledged.

## References

1. Abraham, E.P., Chain, E. An enzyme from bacteria able to destroy penicillin. *Rev. Infect. Dis.* **1988**, *10*, 677–78.
2. Arendrup, M.,C. Update on antifungal resistance in *Aspergillus* and *Candida*. *Clin. Microbio. Inf.*, Special Issue: *Invasive Fungal Infections: Epidemiology and Treatment.* **2014**, *20*, Issue Supplement s6, 42–48.
3. Beach, D. H.; Chen, F.; Cushion, M.T.; Macomber, R. S.; Krudy, G. A.; Wyder, M. A.; Kaneshiro, E. S. Effects of steroidal allenic phosphonic acid derivatives on the parasitic protists *Leishmania donovani*, *Leishmania mexicana mexicana*, and *Pneumocystis carinii carinii*. *Antimicrob. Agents Chemother.* **1997**, *41*, 162–168.
4. Bertrand-Harb, C.; Ivanova, I.; Dalgalarondo, M.; Hartle, T. Evolution of  $\alpha$ -lactoglobulin and  $\beta$ -lactalbumin content during yoghurt fermentation. *Int. Dairy J.* **2003**, *13*, 39–45.
5. Braykov, N. P., Eber, M.R., Klein, E.Y., Morgan, D.J., Laxminarayan, R. Trends in resistance to carbapenems and third-generation cephalosporins among clinical isolates of *Klebsiella pneumoniae* in the United States, 1999–2010. *Infect. Control Hosp. Epidemiol.* **2013**, *34*, 259–68.
6. Brinsley, K, Srinivasan, A, Sinkowitz-Cochran, R, et al. Implementation of the Campaign to Prevent Antimicrobial Resistance in Healthcare Settings: 12 steps to prevent antimicrobial resistance among hospitalized adults-experiences from 3 institutions. *Am. J. Infect. Control* **2005**, *33*, 53–54.
7. Centers for Disease Control and Prevention (CDC). Vital signs: carbapenem-resistant Enterobacteriaceae. *MMWR Morb. Mortal Wkly. Rep.* **2013**, *62*, 165–70.
8. Davies, J, Davie, S D. Origins and evolution of antibiotic resistance. *Microbiol. Mol. Biol. Rev.* **2010**, *74*, 417–33.
9. Goossens, H., Ferech, M., Vander Stichele R., Elseviers M., for the ESAC Project Group. Outpatient antibiotic use in Europe and association with resistance: a cross-national database study. *Lancet* **2005**, *365*, 579–87.
10. Ignatova-Ivanova, TS., Stefanova, I.; Ismailov, I. E.; Ivanov, I. K.; Christov, V. Ch. *In Vitro* Studies of antibacterial activity of a bifunctionalized allene ethanol extracts. *Int. J. Curr. Microbiol. App. Sci.* **2015**, *4*, 589–595.
11. Ignatova-Ivanova, TS., Stefanova, I. ; Ismailov, I. E.; Ivanov, I. K.; Christov, V. Ch. *In Vitro* Studies of antifungal activity of a bifunctionalized allene ethanol extracts. *Int. J. Res. Stud. BioSci. (IJRSB)* **2015**, *3*, 39–40.
12. Ismailov, I. E.; Ivanov, I. K.; Christov, V. Ch. Bifunctionalized allenes. Part XIII. A convenient and efficient method for regioselective synthesis of phosphorylated  $\alpha$ -hydroxyallenes with protected and unprotected hydroxy group. *Molecules* **2014**, *19*, 6309–6329; doi:10.3390/molecules19056309.
13. Klein, E, Smith, D. L, Laxminarayan, R. Hospitalizations and deaths caused by methicillin-resistant *Staphylococcus aureus*, United States, 1999–2005. *Emerg. Infect. Dis.* **2007**, *13*, 1840–46.
14. Laxminarayan, R., Heymann, D.L. Challenges of drug resistance in the developing world. *BMJ* **2012**, *344*, e1567.
15. National Committee for Clinical Laboratory Standards (NCCLS), *Performance standards for antimicrobial disc susceptibility tests*. Approved standard NCCLS Publication, Villanova, PA, USA, **1993**, M2–A5.
16. Omura, S.; Pyl, D.V.D., Inokoshi, J., Takahashi, Y., Takeshima, H. Peptidicinnamins, new farnesylprotein transferase inhibitors produced by an actinomycete. I. Producing strain, fermentation, isolation and biological activity. *J. Antibiotics*, **1993**, *46*, 222–228.
17. Ramanan Laxminarayan, Adriano Duse, Chand Wattal, Anita K. M. Zaidi, Heiman F. L. Wertheim, Nithima Sumpradit, Erika Vlieghe, Gabriel Levy Hara, Ian M

- Gould, Herman Goossens, Christina Greko, Anthony D So, Maryam Bigdeli, G ran Tomson, Will Woodhouse, Eva Ombaka, Arturo Quizhpe Peralta, Farah Naz Qamar, Fatima Mir, Sam Kariuki, Zulfi qar A Bhutta, Anthony Coates, Richard Bergstrom, Gerard D Wright, Eric D Brown, Otto Cars. Antibiotic resistance - the need for global solutions. *The Lancet Infectious Diseases Commission*.**2015**,*13*, 1057–98,[http://dx.doi.org/10.1016/S1473-3099\(13\)70318-9](http://dx.doi.org/10.1016/S1473-3099(13)70318-9).
18. Schwaber, M. J., Lev, B., Israeli, A., *et al*, for the Israel Carbapenem-Resistant Enterobacteriaceae Working Group. Containment of a country-wide outbreak of carbapenem-resistant *Klebsiella pneumoniae* in Israeli hospitals via a nationally implemented intervention. *Clin. Infect. Dis.***2011**,*52*,848–55.
19. Stone, S. P., Fuller, C., Savage, J., *et al*. Evaluation of the national Clean your hands campaign to reduce *Staphylococcus aureus* bacteraemia and *Clostridium difficile* infection in hospitals in England and Wales by improved hand hygiene: four year, prospective, ecological, interrupted time series study. *BMJ***2012**,*344*,e3005.
20. World Health Organization. The evolving threat of antimicrobial resistance. Options for action. ISBN 978 92 4 150318 1. Design by GPS PUBLISHING, France, **2012**.

**How to cite this article:**

A Ignatova-Ivanova TS *et al*, Antibacterial Activity of a Bifunctionalized Allene Ethanol extracts. *International Journal of Recent Scientific Research Vol. 6, Issue, 6, pp.4571-4575, June, 2015*

\*\*\*\*\*